

# KAPA PROBE FORCE

Evolved to break through.



KAPA PROBE FORCE is our most inhibitor-resistant qPCR master mix that removes the need for DNA purification, enabling streamlined sample-to-C<sub>q</sub> workflows. The master mix contains a third generation DNA polymerase evolved to overcome blood, tissue, and plant PCR inhibitors. Crude samples can now be analyzed with comparable accuracy, reproducibility, and sensitivity as purified DNA using KAPA PROBE FORCE.

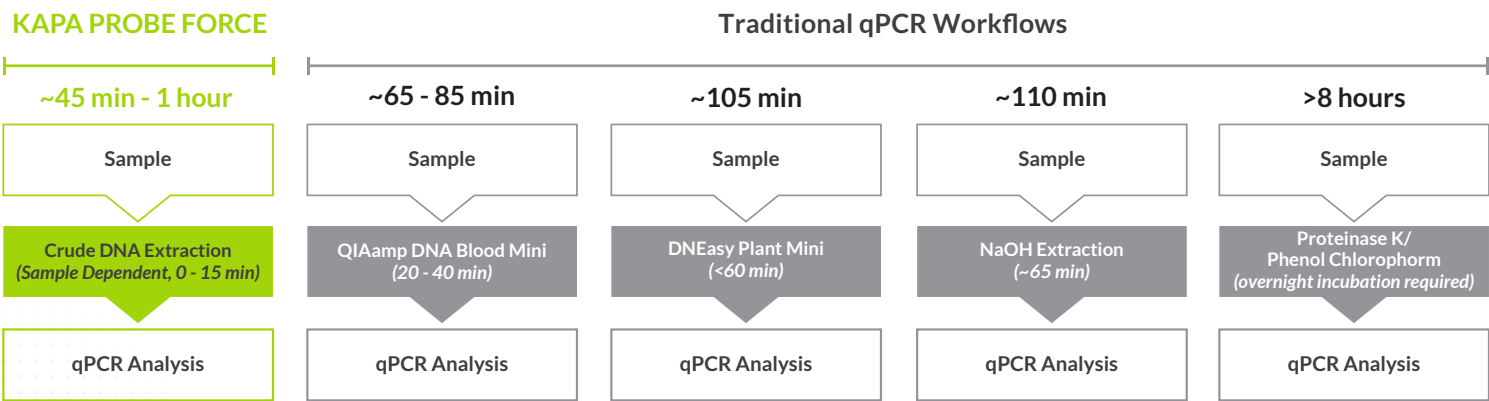
#### Benefits include:

- direct qPCR from crude blood, tissue, and plant extracts
- sample-to-C<sub>q</sub> workflows in <1 hour
- high efficiency for accurate, reproducible, and sensitive results
- superior tolerance to carry-over inhibitors
- multiplex compatibility with crude extracts

# Streamline Sample-to-C<sub>q</sub> Workflows

KAPA PROBE FORCE enables the use of rapid crude DNA extraction methods and overcomes carry-over inhibitors. Competing master mixes used in traditional blood, tissue, and plant qPCR workflows require robust upstream sample processing (e.g. column purification or nuclease digestion).

- Eliminate the time and cost of sample purification by amplifying directly from crude samples
- Analyze a wide range of sample types including whole blood, cells, mouse tails, FFPE, leaf, stem, seed, and soil



## Generate Accurate and Reproducible Results

- Kits include a third-generation DNA polymerase, evolved for robust target amplification and detection
- Enzyme maintains high reaction efficiency in the presence of PCR inhibitors for reliable data generation



**High efficiency target amplification.** Reaction efficiencies achieved for inhibitor spiked samples were examined and compared to that of purified DNA. Across various inhibitor types, efficiencies remained within 90 - 110%.

# Break Through High Levels of qPCR Inhibitors

KAPA PROBE FORCE exhibits consistent and robust amplification across all inhibitors tested, without observable  $C_q$  delays.

- Achieve greater levels of sensitivity for inhibited blood, tissue, and plant samples
- Convert purified DNA assays to crude workflows without observable  $C_q$  delays

*Purified vs. Inhibited Sample  $\Delta C_q$*

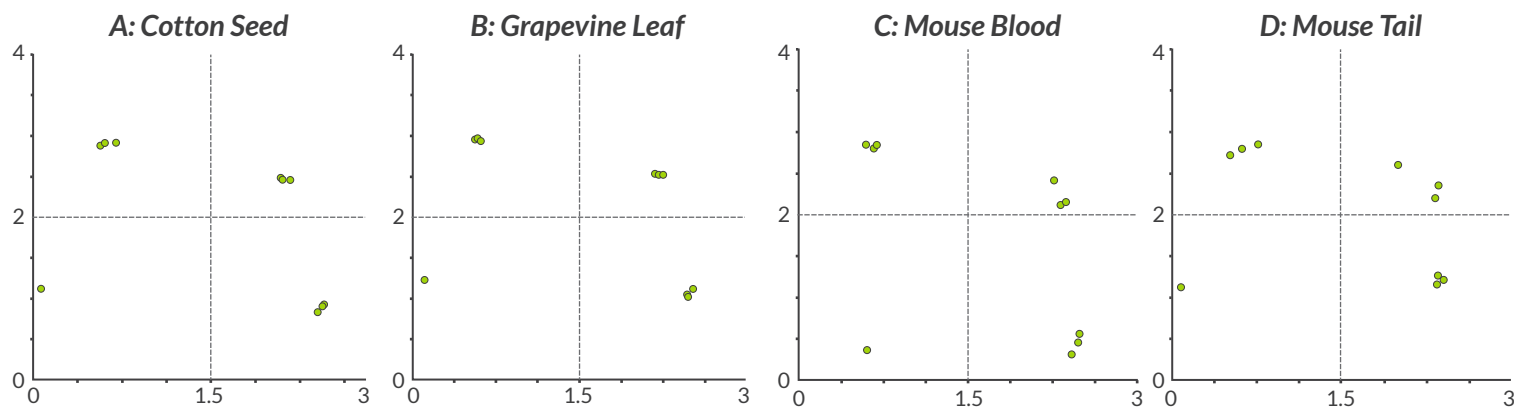
|                                    |                                    | PROBE FORCE | Competitor 1 | Competitor 2 | Competitor 3 | Competitor 4 | Competitor 5 |
|------------------------------------|------------------------------------|-------------|--------------|--------------|--------------|--------------|--------------|
| 100 pg human gDNA                  |                                    | 29.62       | 28.91        | 29.08        | 32.98        | 29.53        | 29.78        |
| Blood Inhibitors                   | Citrate (3.5 mM)                   | -0.04       | 2.64         | -0.18        | 0.98         | 0.20         | 2.90         |
|                                    | EDTA (2 mM)                        | 0.26        | 0.29         | 0.24         | -0.35        | 0.80         | 1.07         |
|                                    | Ferritin (9 $\mu$ g /10 $\mu$ L)   | -0.33       | 0.50         | 0.48         | 10 ng        | NA           | NA           |
|                                    | Hematin (100 $\mu$ M)              | 0.99        | 0.29         | 0.75         | NA           | NA           | NA           |
|                                    | Heparin (750 pg /10 $\mu$ L)       | -0.23       | 0.67         | 1.14         | -0.02        | 0.53         | 3.77         |
| 100 pg mouse gDNA                  |                                    | 29.56       | 29.17        | 28.78        | 32.40        | 29.13        | 29.15        |
| Tissue Inhibitors                  | Collagen (200 ng /10 $\mu$ L)      | -0.41       | 0.63         | -0.02        | 1.40         | 0.21         | 0.69         |
|                                    | Myoglobin (10 $\mu$ g /10 $\mu$ L) | 0.18        | 1.59         | 4.84         | -1.65        | 3.47         | 1.97         |
|                                    | Melanin (50 ng /10 $\mu$ L)        | -0.09       | 0.73         | 0.97         | NA           | NA           | NA           |
|                                    | CaCl <sub>2</sub> (10 mM)          | 0.03        | 100 ng       | 100 ng       | NA           | 100 ng       | NA           |
|                                    | DMEM (30%)                         | -0.72       | NA           | NA           | NA           | NA           | NA           |
| 40 pg grapevine gDNA               |                                    | 33.79       | 33.85        | 33.70        | 34.29        | 33.05        | 40.78        |
| Plant Inhibitors                   | Polyphenols (7%)                   | 1.02        | 0.10         | 0.47         | 3.01         | 0.98         | 1 ng         |
|                                    | Humic Acid (150 ng/10 $\mu$ L)     | 0.76        | 0.52         | 0.70         | NA           | NA           | NA           |
| 20 fg purified <i>E. coli</i> gDNA |                                    | 31.18       | 30.75        | 31.16        | 35.90        | 31.22        | 44.80        |
| Extraction Inhibitors              | Ethanol (3%)                       | -0.03       | 0.56         | -0.41        | 20 pg        | -0.23        | NA           |
|                                    | NaAc (60 mM)                       | 0.42        | 1.27         | 20 pg        | -4.15        | NA           | NA           |
|                                    | NaCl (60 mM)                       | 0.14        | NA           | 200 pg       | 20 pg        | NA           | NA           |

■ <1  $\Delta C_q$ 
■ 1 - 2  $\Delta C_q$ 
■ 2 - 3  $\Delta C_q$ 
■ >3  $\Delta C_q$ 
■ Detection failed. Lowest concentration at which  $C_q$  < 45 cycles detected or No Amplification (NA).

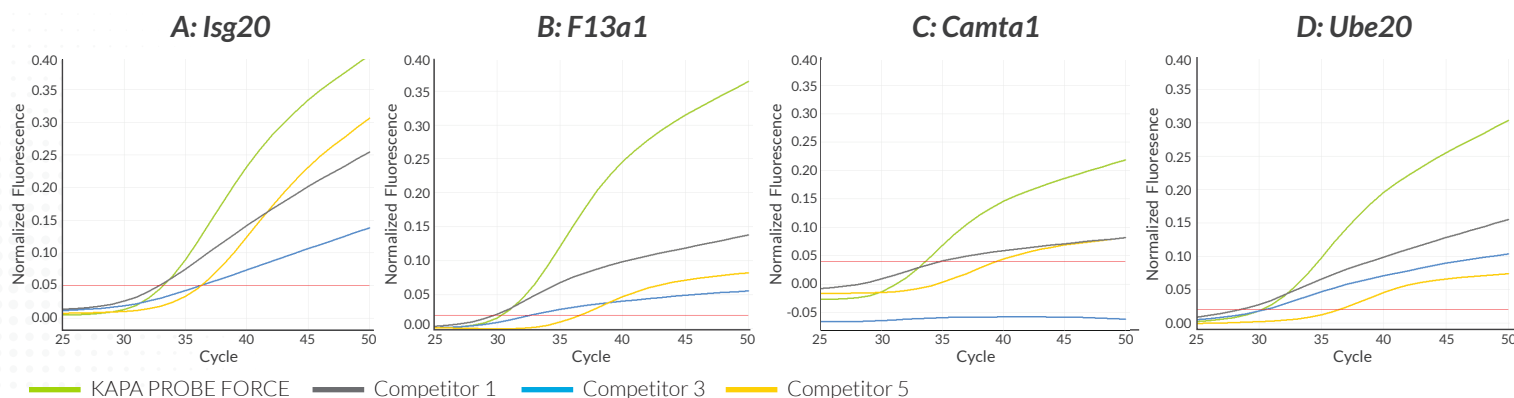
**Broad range of high inhibitor resistance.** Baseline performance of KAPA PROBE FORCE and competing master mixes was measured by creating standard curves with purified DNA according to each manufacturer's recommended cycling conditions. Serial dilutions were run in the following ranges: Human: 100 ng – 10 pg; Mouse: 100 ng – 10 pg; Plant: 25 ng – 8 pg; and Bacteria: 2 ng – 2 fg. Inhibitors were individually spiked into purified DNA samples at high concentrations to determine their effect on  $C_q$  values.

# Multiplex Crude Samples Efficiently

- Accelerate genotyping analysis with single reaction allelic discrimination of crude DNA extracts
- Maximize data collection from precious samples, increase throughput, and reduce costs



**Crude sample duplex SNP detection.** KAPA PROBE FORCE provides accurate genotyping and tight clustering in the presence of crude extracts (A) cotton seed, 0.5M NaOH extraction; (B) grapevine leaf, 75 mM Tris-HCl and 5 mM TCEP extraction; (C) mouse blood, FTA 0.5 mm disc; and (D) mouse tail extracts, NaOH extraction, for rapid SNP analysis.



**Highly efficient 4-plex performance.** Four targets were amplified in a multiplex assay with KAPA PROBE FORCE and three competitive master mixes. 100 pg mouse gDNA was amplified targeting the (A) Isg20 (FAM/BHQ-1), (B) F13a1 (CAL Fluor Orange 560), (C) Camta1 (Quasar 670) and (D) Ube20 (Quasar 705) genes. 500 nM primers and 110 nM probes were used with the following cycling conditions: 95°C for 30 sec followed by 50 cycles of 95°C for 3 sec, and 60°C for 30 sec.

## Ordering Information

| Kit Code | Description                                     | Kit Size |
|----------|---|----------|
| KK4300   | KAPA PROBE FORCE qPCR Master Mix (2X) Universal | 1 mL     |
| KK4301   | KAPA PROBE FORCE qPCR Master Mix (2X) Universal | 5 mL     |
| KK4302   | KAPA PROBE FORCE qPCR Master Mix (2X) Universal | 10 mL    |
| KK4303   | KAPA PROBE FORCE qPCR Master Mix (2X) Universal | 50 mL    |

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