



# Rapid Prep for PCR Applications Protocol 2

## Lyse & Purify (16 Tubes) Cell Suspensions and Cell Pellets

Rapidly lyse and purify DNA from cell suspensions or cell pellets. This kit contains materials for processing 16 samples.

### **Additional Items Needed: (Not included; see Suggested Supplier list below)**

1. **Additional Items Required** for Lyse and Purify
  - a. 8-strip 0.2 mL PCR tubes w/individual domed caps for sample lysis and purification
  - b. Sterile Laboratory Grade Water
  - c. Fresh 70% ethanol
  - d. TE Buffer - 10mM Tris, 1mM EDTA, pH 8.0
  - e. 0.4M KOH Solution
  - f. Magnetic Rack ('bar' or 'post' suggested for higher DNA yield)
2. **Suggested items for rapid processing**
  - a. Repeater Pipette with multi-volume tips
  - b. Single Channel Manual Pipettes
  - c. Multichannel Pipette 0.5-10  $\mu$ L working volume
  - d. Mini Centrifuge for 200  $\mu$ L 8-strip tubes

### **Kit Contents for Shoreline Biome Lyse and Purify**

1. Two 8-tube strips of dried Lysis-1
2. Purification Buffer (two 1.5 mL tubes with **green cap**)

### **Shoreline Biome Lyse and Purify Protocol**

1. **Make 0.4M KOH.** Measure 0.135g KOH into 15 mL tube, then add 6 mL H<sub>2</sub>O. 0.4M KOH expires after 7 days.
2. **Make 70% Ethanol.** Add 17.5 mL 100% ethanol to 7.5 mL sterile laboratory grade water
3. Bring Purification Buffer (**green capped tube**) to room temperature.
4. **Label a clean 200  $\mu$ L tube for each sample** (e.g., for 16 samples, label two 8-strip 200  $\mu$ L tubes)
5. **Sample Type:**
  - 5.1. **Cell suspension:**
    - 5.1.1. **Transfer 50  $\mu$ L of cell suspension** (between 10<sup>6</sup> and 10<sup>8</sup> cells) to Lysis-1 tube. Do not mix.
  - 5.2. **Cell pellets:**
    - 5.2.1. Reconstitute the cell pellet by adding 50  $\mu$ L Molecular Biology Grade water, TE, or PBS and pipetting up and down to suspend cells.
    - 5.2.2. **Transfer 50  $\mu$ L** of the cell suspension (between 10<sup>6</sup> and 10<sup>8</sup> cells) to Lysis-1 tube. Do not mix.
6. **Add 50  $\mu$ L 0.4M KOH** to each tube. Do not mix. (A precipitate will form)
7. Cap samples tightly; set PCR machine for proper tube height, close and lock the lid and **heat samples to 95°C for 5 minutes.** Precipitate will dissolve. After incubation, **allow sample to cool to RT before unlocking PCR lid** – failure to do so may result in lids popping off tubes.
8. **While samples are cooling:** Vortex Purification Buffer tubes (**Green Cap**) to re-suspend any brown magnetic beads that may have settled
9. Add **100  $\mu$ L of Purification Buffer** (**Green Cap**) to each sample. If precipitate formed, it should re-dissolve. Pipette up and down to mix.



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10. Cap samples tightly; set PCR machine for proper tube height, close and lock the lid, and **heat samples to 50°C for 3 minutes** to allow DNA to bind to beads.
11. **After 50°C incubation, place samples on magnetic rack** to pellet beads with bound DNA to side of the tube (~60 seconds)
12. Once all beads are pelleted, **remove all supernatant SLOWLY and discard**. Avoid aspirating beads.
13. **Remove tubes from magnet, add 100 µL 70% ethanol** (at room temperature) to each tube, and re-suspend beads by pipetting gently up and down.
14. **Return tubes to magnet and allow beads to pellet** (~30 seconds)
15. **Pipette off residual ethanol and let air dry for 2 - 3 minutes**. Over-drying will make eluting DNA more difficult, so watch to ensure ethanol is mostly gone but bead pellet does not begin to shrink and crack.
16. **Remove tubes from magnet**.
17. Add **20 µL of 1X TE buffer** to tubes and briefly pipette up and down to re-suspend beads. Solution should turn brown in color to reflect beads mixed homogeneously throughout.
18. **Let solution incubate at room temperature for 2 minutes**.
19. **Place tubes on magnetic rack** for 30 - 60 seconds to pellet magnetic beads. **DNA is now in solution.**
20. **Transfer supernatant containing eluted DNA** into **clean**, labeled microcentrifuge tubes on ice, avoiding pelleted beads.
21. **Remove tubes with pelleted beads from rack**.
22. **Add another 20 µL 1X TE buffer** onto bead pellets, gently pipette up and down to re-suspend beads.
23. **Incubate tubes for 2 minutes at room temperature**.
24. Return tubes to magnetic rack. Allow beads to pellet for 30 - 60 seconds.
25. **Remove supernatant containing DNA** and combine with first eluate in clean, labeled tubes, avoiding pelleted beads.
26. **Store eluted DNA at +4°C**.

**NOTES:** DNA is **denatured**, due to KOH treatment during lysis, which is optimal for PCR. Store Lysis Mix at +4°C for up to 10 days, or frozen at -20°C for 3 months. Avoid repeated freeze/thaws.

### **Suggested Suppliers**

Dot Scientific Inc.: 8-strip PCR tubes w/individual domed caps, #503-8PCR

Fisher Scientific: Biotech Grade Water BP24854; Potassium Hydroxide P250; TE BP24731

Permagen: 0.2 mL PCR Strip Magnetic Separation Rack 8 or 12 Strip, Cat #MSR812

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