

CHOgro® Transfection and Titer Enhancer Kit

Quick Reference Protocol

Instructions for MIR 6225

Full protocol, SDS and Certificate of Analysis available at [mirusbio.com/6225](https://mirusbio.com/6225)



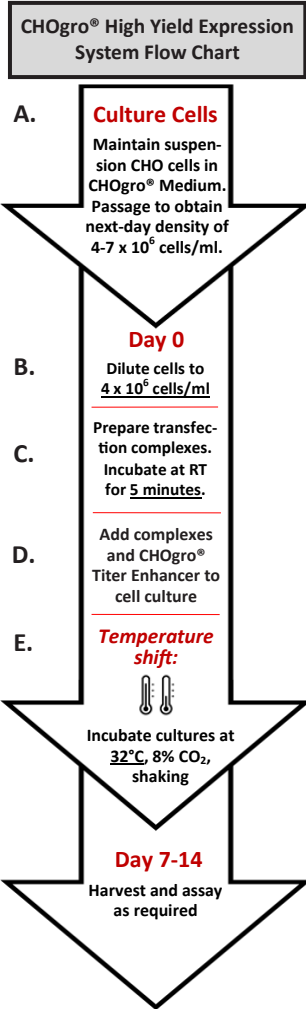
SPECIFICATIONS

Storage	Store <i>TransIT</i> -PRO® Transfection Reagent (MIR 5740) tightly capped at -20°C. Store CHOgro® Titer Enhancer (MIR 6220) at 2-10°C, protected from light. <i>Before each use</i> , warm to room temperature and vortex gently.
Product Guarantee	1 year from the date of purchase, when properly stored and handled.
Intended Usage	Designed for use with CHOgro® High Yield Expression System (MIR 6270)

► PLASMID DNA TRANSFECTION PROTOCOL



Full protocol and additional documentation available at [mirusbio.com/6225](https://mirusbio.com/6225)



Fill in volumes below based on total culture volume (Table 1).

- A. Maintenance of cells**
1. Passage cells 18–24 hours prior to transfection to obtain a next day density of 4-7 x 10<sup>6</sup> cells/ml. DO NOT proceed with transfection if cells are not doubling every 24 hours or are < 98% viable.
  2. Incubate cells overnight at 37°C in 8% CO<sub>2</sub>, shaking.
- B. Prepare/dilute suspension CHO cells for transfection**
1. Seed cells at 4 x 10<sup>6</sup> cells/ml immediately before transfection. DO NOT proceed if cells are not doubling normally or < 98% viable.
- C. Prepare *TransIT*-PRO® Reagent:DNA complexes**
1. Warm *TransIT*-PRO® Reagent to room temperature. Vortex gently.
  2. Place \_\_\_\_ ml CHOgro® Complex Formation Solution in sterile tube.
  3. Add \_\_\_\_ µg plasmid DNA. Mix gently by pipetting.
  4. Add \_\_\_\_ µl of *TransIT*-PRO® Reagent. Mix gently by pipetting.
  5. Incubate at room temperature for no more than 5 minutes to allow transfection complexes to form.
- D. Add transfection complexes and CHOgro® Titer Enhancer to cells**
1. Add transfection complexes to cell culture. Mix gently by swirling.
  2. Add \_\_\_\_ µl of CHOgro® Titer Enhancer to cell culture + transfection complexes. Mix gently by swirling.
- E. Place culture at 32°C after transfection and enhancer addition**
1. Incubate cultures at 32°C (8% CO<sub>2</sub>, shaking) for 2-14 days.  
NOTE: Optimal culture time will depend on cell type, protein of interest, culture temperature, and detection method.
  2. Harvest cells and/or supernatant and assay as required.

Table 1. Scaling worksheet for CHOgro® Transfection and Titer Enhancer Kit.

Starting transfection conditions per milliliter of CHOgro® Expression Medium:				
	Per 1 ml		Total culture volume	Reagent quantities
Complex Formation Solution	0.1 ml	x	____ ml	= ____ ml
Plasmid DNA (1 µg/µl stock)	1 µl	x	____ ml	= ____ µl
<i>TransIT</i> -PRO® Reagent	1 µl	x	____ ml	= ____ µl
Enhancer addition (add to culture after transfection complex addition):				
CHOgro® Titer Enhancer	20 µl	x	____ ml	= ____ µl

## ► Critical Parameters for Success with CHOgro® High Yield Expression System

- **Cell adaptation and maintenance.** Cells grown in alternate media formulations should be fully adapted to CHOgro® Expression Medium supplemented with 4mM L-Glutamine and 0.3% Poloxamer 188 prior to transfection with the CHOgro® High Yield Expression System. Cells are fully adapted when they are ≥98% viable and doubling normally.
- **Cell density at transfection.** Cells should be passaged 18–24 hours prior to transfection to obtain a next day density of  $4\text{--}7 \times 10^6$  cells/ml. This allows for a minimal cell dilution for a final density of  $4 \times 10^6$  cells/ml at the time of transfection. Cultures should be placed at 37°C in 8% CO<sub>2</sub> prior to transfection. DO NOT proceed with transfection if cells are not doubling daily and at least 98% viable by trypan blue exclusion.
- **DNA concentration.** Start with 1 µg of DNA per 1 ml of culture. Vary the DNA concentration from 1–2 µg/ml to find the best working DNA concentration. To maintain the *TransIT*-PRO® Reagent:DNA ratio, adjust the reagent volume accordingly. NOTE: Use only high quality, endotoxin-free DNA for transfections. Contaminants such as protein, carbohydrate and lipids may affect transfection efficiency and gene expression levels. Ensure that the plasmid preparation exhibits an A260/A280 ratio of > 1.8.
- **Ratio of *TransIT*-PRO® Reagent to DNA.** Start with 1 µl of *TransIT*-PRO® Reagent per 1 µg of DNA. Vary the concentration of *TransIT*-PRO® Reagent from 1–2 µl per 1 µg of DNA to find the optimal ratio.
- **Transfection complex formation.** Prepare *TransIT*-PRO® Reagent:DNA complexes in CHOgro® Complex Formation Solution (MIR 6210). Incubate complexes at room temperature for no more than 5 minutes before adding to cultures directly.
- **CHOgro® Titer Enhancer addition.** CHOgro® Titer Enhancer should be added to the culture immediately after transfection complex addition. Add 20 µl of CHOgro® Titer Enhancer per 1 ml cell culture (see Table 1 on front page for scaling chart). Cultures should then be placed at 32°C, 8% CO<sub>2</sub> (shaking) for the remainder of the culture.
- **Feeds.** No feeds are required for high yield, but an optional feed can be added to prolong cellular viability (see [CHOgro® High Yield Expression protocol](#) for details).
- **Temperature shift to 32°C post-transfection.** Placing flasks at 32°C immediately post-transfection will increase overall protein titers and decrease protein degradation. Typically, greater than 2-fold higher antibody titers are achieved if incorporating the temperature shift into the production workflow.
- **Post-transfection incubation time.** The optimal post-transfection incubation time may vary by the experimental goal and the plasmid used. For secreted antibody constructs, optimal titers are obtained at 32°C at 7–14 days post-transfection in batch culture.



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